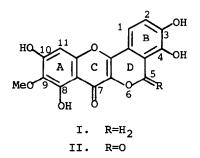
Tetrahedron Letters No. 46, pp 4507 - 4510. ©Pergamon Press Ltd. 1979. Printed in Great Britain. 0040-4039/79/1108-4507\$02.00/0

STRUCTURE AND SYNTHESIS OF A DERIVATIVE OF FASCICULIFERIN, A NOVEL PELTOGYNOID FROM ACACIA FASCICULIFERA

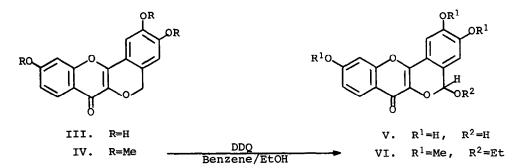
Fanie R. van Heerden, E. Vincent Brandt and David G. Roux^{*} Department of Chemistry, University of the Orange Free State, P.O. Box 339, Bloemfontein 9300, South Africa

The structure and synthesis of the 5-0-ethyl-2,3-10-tri-0-methyl derivative of fasciculiferin (5-hydroxypeltogynin) a natural peltogynoid which exhibits an intermediate oxidation state of the C-5 methylene function in the D-ring are described.

Hitherto only two natural variants of the oxidation state of the 'extra' D-ring carbon of C_{16} -peltogynoids have been recorded. These are illustrated by, and indeed limited to, the pair of flavonol analogues, benthamianin (I) with C-5 in the reduced form, and distemonanthin (II), with it fully oxidized; com= pounds which are associated in the heartwood of *Distemonanthus benthamianus*.^{1,2,3}



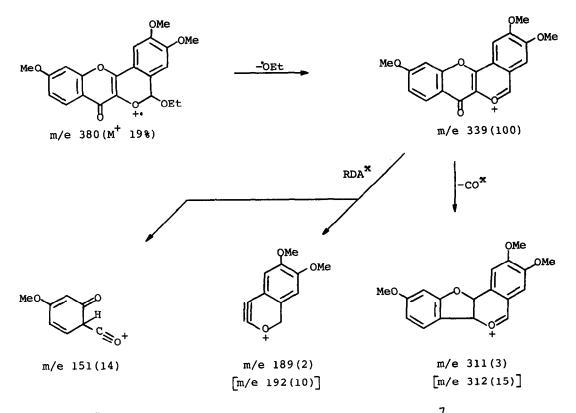
Indirect evidence is now presented for the natural existence in another source of the first example of a peltogynoid with a C-5, or corresponding D-ring, in an intermediate oxidation state. The heartwood of Acacia fasciculifera F. Muell ex Benth. contains amongst related flavonoid metabolites, also (+)-2,3-trans-3,4-trans-peltogynol, pelto-gynin (III), 4,5 and a novel peltogynoid (R_F 0.0 in 2% HOAC, and ~0.7 in watersatd. butan-2-ol), all in low proportion. The latter was isolated in 0.2% yield as racemic 5-ethoxy-2,3-10-trimethoxypeltogynin (VI), m.p. 160^o (amorph.), following preparative paper chromatography in 2% acetic acid, stripping with 80% ethanol, and methylation with diazomethane.



The structure of the trimethyl ether of 5-ethoxypeltogynin (VI), $C_{21}H_{20}O_7$, [R_F 0.40 in benzene-acetone 4:1 on Kieselgel PF₂₅₄, followed by separation (2x), R_F 0.13 in benzene-EtOAc 7:3; yellow with H₂SO₄-HCHO] is consistent with its n.m.r. [δ (CDCl₃) 8.10(d, J 7.5, H-8), 7.28(s, H-1), 6.88(d, J 2.0, H-11), 6.85 (dd, J 7.5 and 2, H-9), 6.76(s, H-4), 6.03(s, H-5), 3.87,3.92,3.97(s, 3 x OMe), 3.81(q, J 7.5, O<u>CH₂CH₃</u>) and 1.19(t, J 7.5, OCH₂<u>CH₃</u>)] and mass fragmentation spectra (*cf.* scheme). Chemical shift of the 5-proton (δ 6.03) is also in close agreement with that of 5-methoxy- β -photomethylquercetin (δ 5.93)⁶ in the same solvent (CDCl₃).

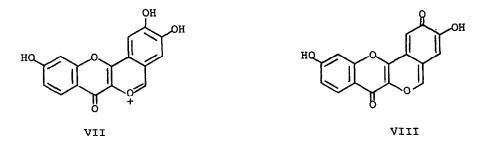
Proof of structure of the derivative was obtained by selective function= alization of the methylene group at C-5 of 2,3,10-tri-0-methylpeltogynin (V)⁵ with DDQ in benzene-ethanol (50:1 v/v), according to a variation of the method of Buchan *et al.*,⁸ to give a 30% yield of the desired product (VI) in a single step conversion (*cf.* scheme).

The 5-ethoxy compound derived from A. fasciculifera is considered to be an



*Presumably accompanied by H-transfer (cf. ref. ⁷)

artefact which originates from 5-hydroxypeltogynin (V) *via* a stable oxonium ion (VII), formed under the acid conditions applied during preparative paper chromatography or during subsequent stripping and by its solvolysis with etha: nol under the latter conditions (80% ethanol:residual acetic acid). Such



presumption for the natural presence of 5-hydroxypeltogynin, is supported by

the observation that oxidative synthesis of its tri-0-methyl ether from 2,3,10-tri-0-methylpeltogynin (IV) is also accompanied by spontaneous reaction with ethanol; formation of an intermediate quinone methide (cf. ref.⁸) which is trapped by ethanol being excluded in this instance.

The susceptibility of the C-5 methylene function in peltogynoids to oxidation was demonstrated by Waiss *et al.*⁶ under conditions of oxidative photolysis of penta-0-methylquercetin in methanol when 5-methoxy-0-photomethyl= quercetin, an analogue of V, is formed albeit in very low yield. Accordingly fasciculiferin (V) is speculatively considered to have its biogenetic origin in a quinone methide intermediate (VIII) originating from peltogynin (III) *via* an oxidative mechanism.

One of us (F.R.v.H.) acknowledges tenure of the Konrad Taeuber Memorial Fellowship (1979) and receipt of the C.S.I.R. Merit Award (1977-1979). Thanks are also due to the Sentrale Navorsingsfonds of this University for financial support, and to Dr. Mary D. Tindale, Royal Botanic Gardens and National Herba= rium, Sydney for collection and authentication of material.

REFERENCES

- 1. F.E. King, T.J. King and P.J. Stokes, J. Chem. Soc., 1954, 4587.
- 2. E. Malan, J.P. Engelbrecht and D.G. Roux, S. Afr: J. Sci., 1974, 70, 21.
- 3. E. Malan and D.G. Roux, J. Chem. Soc. Perkin I, 1979 (in the press).
- 4. S.E. Drewes and D.G. Roux, J. Chem. Soc. (C), 1967, 1407.
- 5. D. Ferreira, J.P. van der Merwe and D. Ferreira, J. Chem. Soc. Perkin I, 1974, 1492.
- A.C. Waiss, R.E. Lundin, A. Lee and J. Corse, J. Amer. Chem. Soc., 1967, 89, 6213.
- 7. S.E. Drewes and H. Budzikiewicz, J. Chem. Soc. (C), 1969, 63.
- 8. G.M. Buchan, J.W.A. Findlay and A.B. Turner, J.C.S. Chem. Comm., 1975, 126.

(Received in UK 5 September 1979)